

Analysis of Proteomics Data using MALDIquant

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Predicting disease



Proteomics

Proteomics

- Study of the entirety of proteins produced by an organism.
- Foci: identification, structure determination, biomarker, pathways, expression.

Mass Spectrometry

Ion Source: MALDI

Matrix-Assisted Laser
Desorption/Ionization

Mass Analyzer: TOF

Time Of Flight ($t \propto \sqrt{\frac{m}{q}}$)

Detector

Quantity
Measurement

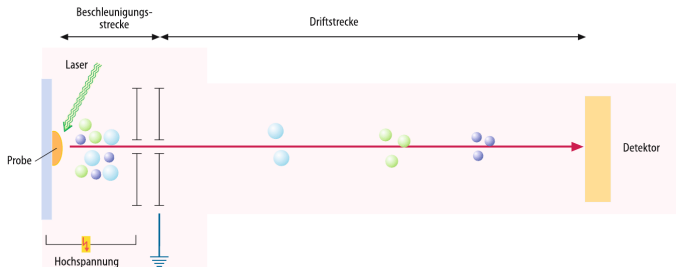
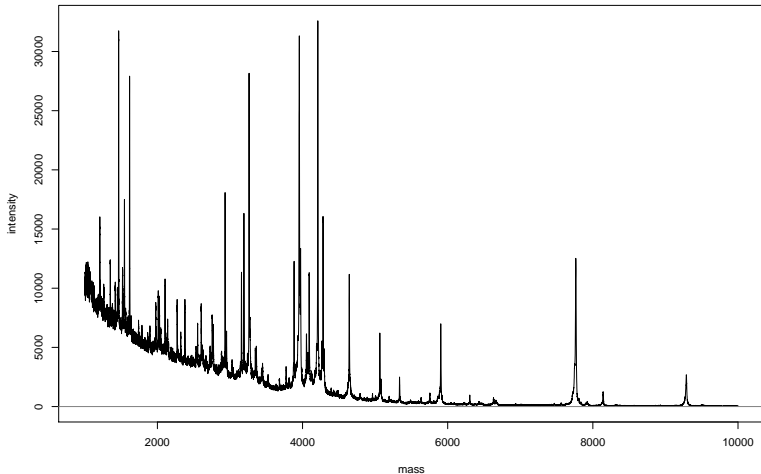


Abb. 3.14.; S. 67; "Biochemie & Pathobiochemie", Löffler G., 8. Auflage (2007), Springer Medizin Verlag

MALDI-TOF Example Spectrum



MALDIquant

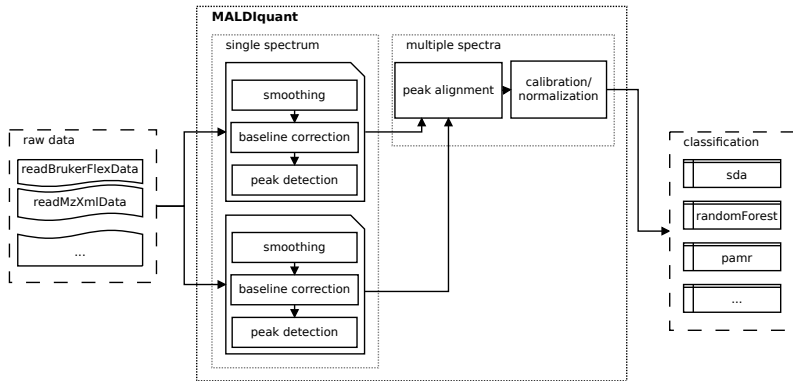
Motivation

- Only relatively few open source software solutions available and very few for the R platform.
- No MALDI-TOF package fitting our needs for clinical diagnostics.
- Necessity of handling both technical and biological replicates.
- Unsatisfying quantification of relative intensities (total-ion-current, 0/1)
- Investigation of impact of calibration of spectra on clinical prognosis.
- Modular and easy to customize analysis routines.

MALDIquant

The UNIX philosophy: do one thing and do it well

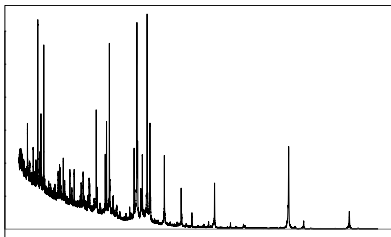
McIlroy (1978)



MALDIquant

The UNIX philosophy: do one thing and do it well

McIlroy (1978)

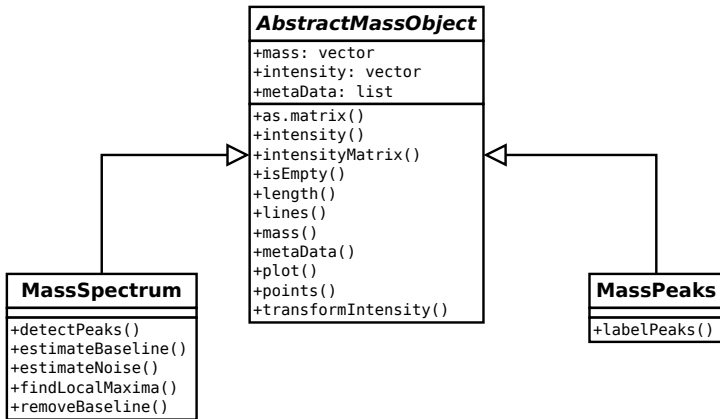


\Rightarrow

$$\begin{pmatrix} f_{1,1} & f_{1,2} & f_{1,3} & \dots & f_{1,n} \\ f_{2,1} & f_{2,2} & f_{2,3} & \dots & f_{2,n} \\ \dots & \dots & \dots & \dots & \dots \\ f_{m,1} & f_{m,2} & f_{m,3} & \dots & f_{m,n} \end{pmatrix}$$

MALDIquant Structure

object-oriented, S4



Example Data

Serum Peptidome Profiling Revealed Platelet Factor 4 as a Potential Discriminating Peptide Associated with Pancreatic Cancer

G.M. Fiedler, A.B. Leichtle, J. Kase et al
Clin Cancer Res June 1, 2009 15:3812-3819

*“Two significant peaks (**m/z 3884**; 5959) achieved a sensitivity of 86.3% and a specificity of 97.6% for the discrimination of patients and healthy controls . . .”*

“MALDI-TOF MS-based serum peptidome profiling allowed the discovery and validation of platelet factor 4 [m/z 3884, 7767; S.G.] as a new discriminating marker in pancreatic cancer.”

MALDIquant

hands-on: File Import

```
> library("MALDIquant")
> library("readBrukerFlexData")
> spectra <- mqReadBrukerFlex("/data/fiedler2009/")
> length(spectra)
[1] 480
> spectra[[1]]
S4 class type           : MassSpectrum
Number of m/z values   : 42388
Range of m/z values    : 1000.015 - 9999.734
Range of intensity values: 5 - 101840
File                   : /data/fiedler2009/[...]/Pankreas_HB_L_061019_G10/0_m19/1/1SLin/fid
```

other possibilities:

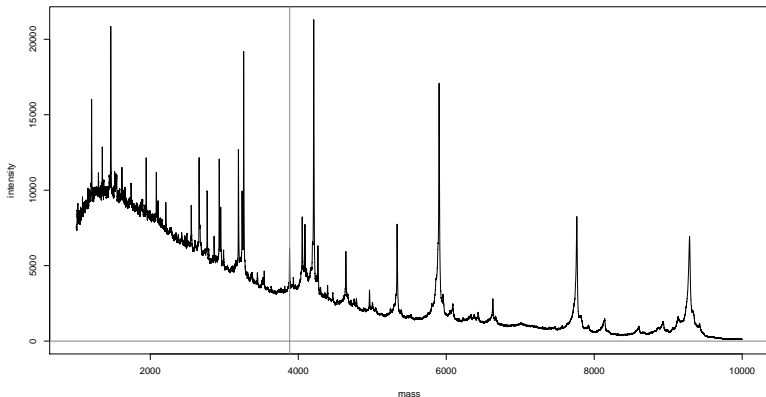
```
library("MALDIquant")
library("readMzXmlData")
s <- mqReadMzXml("/data/exampleMS/spectrum.mzXML")
```

```
library("MALDIquant")
s <- createMassSpectrum(mass=1:5,
                        intensity=runif(5),
                        metaData=list(name="example"))
```

MALDIquant

hands-on: plot

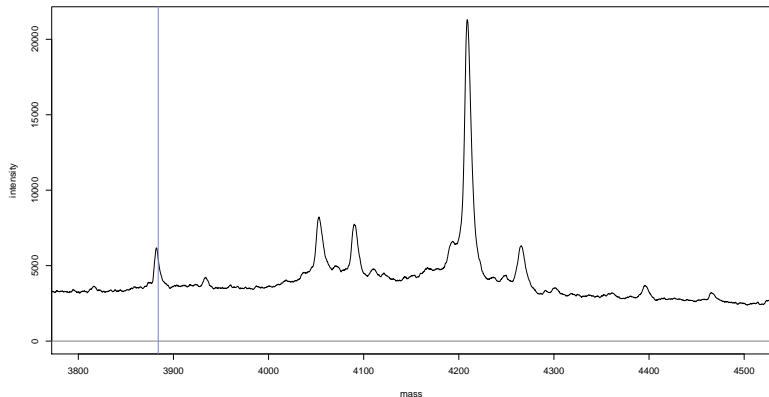
```
> plot(spectra[[1]])  
> abline(v=3884, col="blue")
```



MALDIquant

hands-on: plot m/z 3884

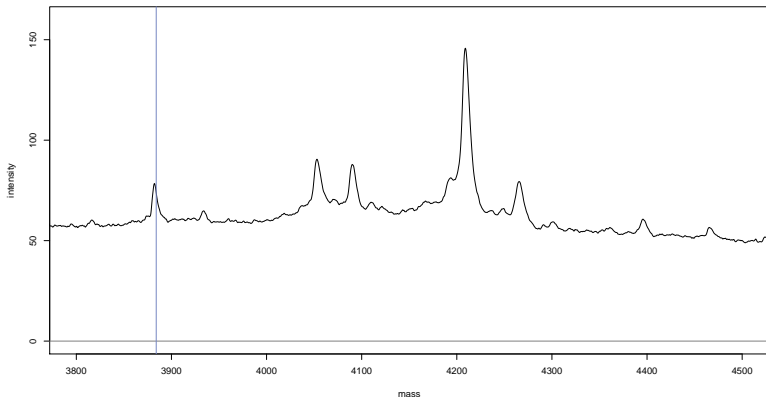
```
> plot(spectra[[1]], xlim=c(3800, 4500))  
> abline(v=3884, col="blue")
```



MALDIquant

hands-on: Variance Stabilization and Smoothing

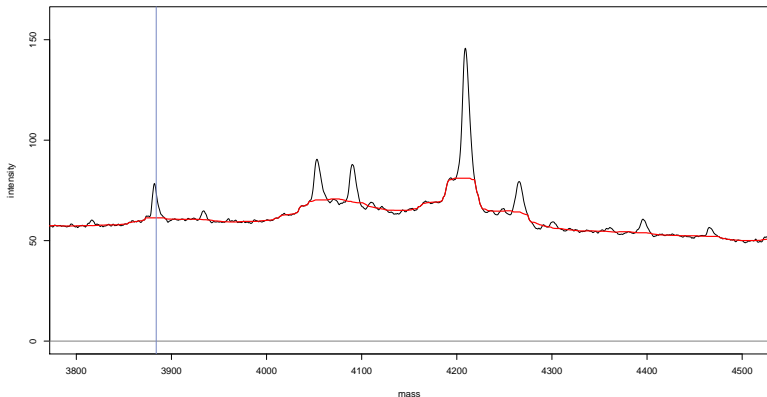
```
> spectra <- lapply(spectra, transformIntensity, fun=sqrt)
> movAvg <- function(y) {return(filter(y, rep(1, 5)/5, sides=2));}
> spectra <- lapply(spectra, transformIntensity, fun=movAvg)
```



MALDIquant

hands-on: Baseline Correction – don't harm the data

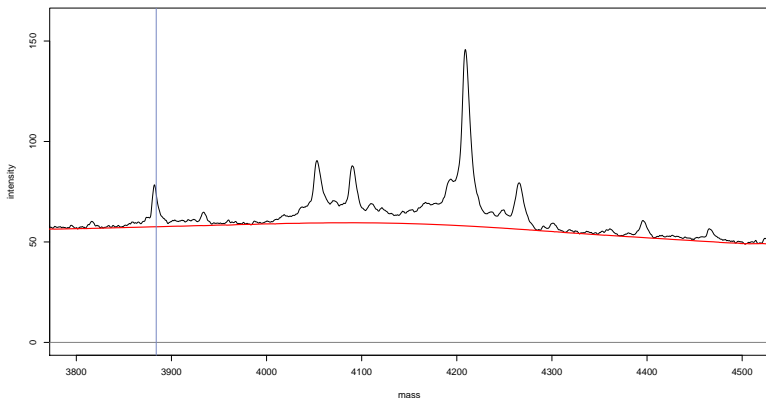
```
> bl <- estimateBaseline(spectra[[1]], method="Median")  
> plot(spectra[[1]], xlim=c(3800, 4500));  
> lines(bl, col="red");
```



MALDIquant

hands-on: Baseline Correction – SNIP

```
> bl <- estimateBaseline(spectra[[1]], method="SNIP")  
> plot(spectra[[1]], xlim=c(3800, 4500)); lines(bl, col="red");
```

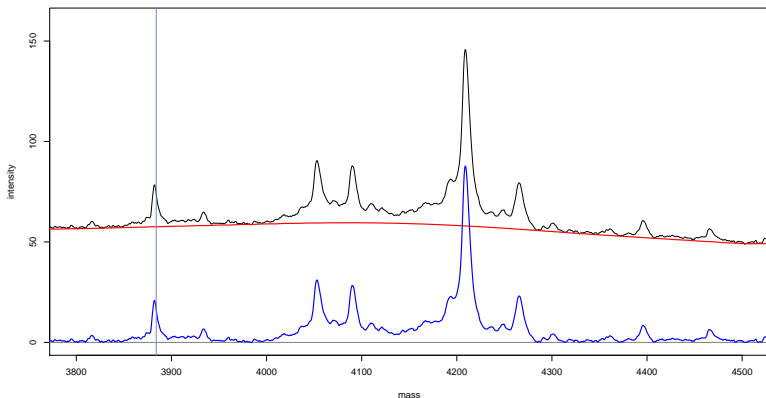


C. G. Ryan, E. Clayton, W. L. Griffin, S. H. Sie, and D. R. Cousens. SNIP, a statistics-sensitive background treatment for the quantitative analysis of PIXE spectra in geoscience applications. *Nucl. Instrument. Meth. B*, 34:396–402, 1988

MALDIquant

hands-on: Baseline Correction – SNIP

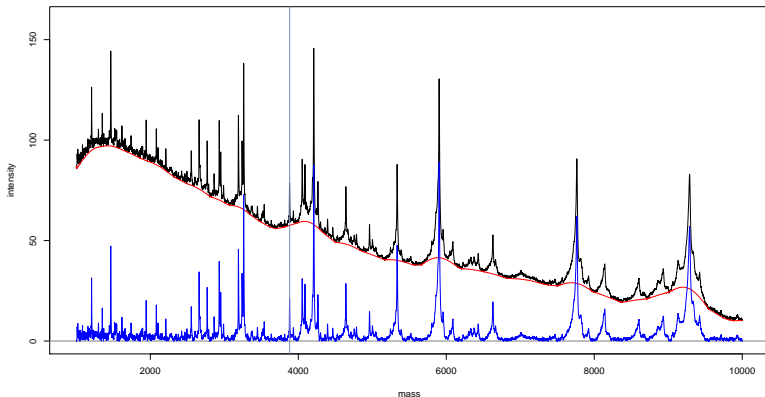
```
> spectra <- lapply(spectra, removeBaseline)
> lines(spectra[[1]], col="blue")
```



C. G. Ryan, E. Clayton, W. L. Griffin, S. H. Sie, and D. R. Cousens. SNIP, a statistics-sensitive background treatment for the quantitative analysis of PIXE spectra in geoscience applications. *Nucl. Instrument. Meth. B*, 34:396–402, 1988

MALDIquant

hands-on: Baseline Correction – SNIP



MALDIquant

hands-on: Peak Detection

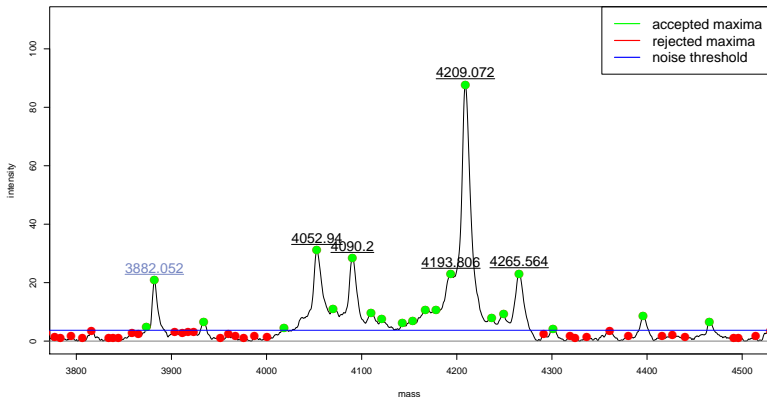
```
> spectra[[1]]
S4 class type      : MassSpectrum
Number of m/z values : 42388
Range of m/z values : 1000.015 - 9999.734
Range of intensity values: 0 - 709.207
File               : /data/fiedler2009/[...]/Pankreas_HB_L_061019_G10/0_m19/1/1SLin/fid
```

```
> peaks <- lapply(spectra, detectPeaks)
> peaks[[1]]
S4 class type      : MassPeaks
Number of m/z values : 198
Range of m/z values : 1011.059 - 9423.422
Range of intensity values: 19.273 - 709.207
File               : /data/fiedler2009/[...]/Pankreas_HB_L_061019_G10/0_m19/1/1SLin/fid
```

MALDIquant

hands-on: Peak Detection

```
> plot(spectra[[1]], xlim=c(3800, 4500)); points(peaks[[1]], col="green")  
> top5 <- intensity(p) %in% sort(intensity(p)[mass(p)>3800 & mass(p)<4500], decreasing=TRUE)[1:5]  
> labelPeaks(peaks[[1]], index=top5); labelPeaks(peak[[1]], mass=3884, col="blue")
```



MALDIquant

Single Spectrum Workflow

```
> library("MALDIquant")
> library("readBrukerFlexData")
> spectra <- mqReadBrukerFlex("/data/fiedler2009/")

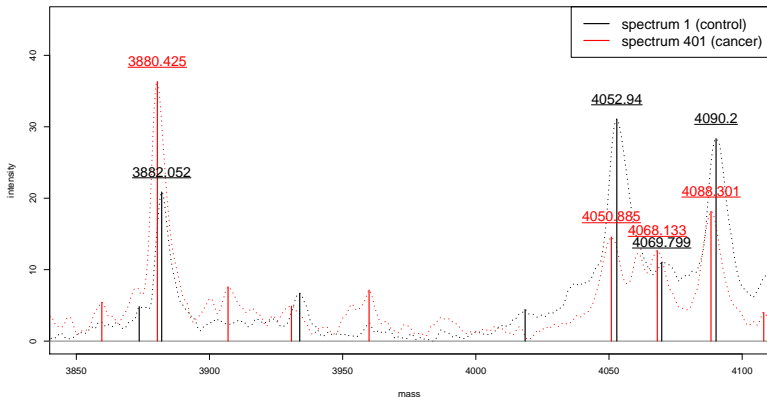
> movAvg <- function(y) {return(filter(y, rep(1, 5)/5, sides=2));}

> spectra <- lapply(spectra, transformIntensity, fun=sqrt)
> spectra <- lapply(spectra, transformIntensity, fun=movAvg)

> spectra <- lapply(spectra, removeBaseline)
> peaks <- lapply(spectra, detectPeaks)
```

MALDIquant

Comparison of Multiple Spectra



MALDIquant

Conclusion

MALDIquant is free software (GPLv3).

Currently available:

- Easy to use and full established single spectrum workflow.
- Easy to customize.
- Easy data exchange.

Shortly available:

- Peak alignment.
- Calibration/Normalization routines.

MALDIquant

Thanks

Alexander B. Leichtle, helpful discussions
(Institute for Clinical Chemistry, Bern University Hospital)

Korbinian Strimmer, supervision
(IMISE, University of Leipzig)

Thanks for your attention!

Download of MALDIquant software:

<http://strimmerlab.org/software/maldiquant/>